



Identification of Immunogenic T and B-Cell Epitope Peptides of Rubella Virus E1 Glycoprotein towards the Development of Highly Specific Immunoassays and Vaccine

Prashanth Rajendiran^a, Nithiyandanan Saravanan^a,
Mageshbabu Ramamurthy^{a*}, Rajasekar Aruliah^b and Balaji Nandagopal^a

^a Sri Sakthi Amma Institute of Biomedical Research, Sri Narayani Hospital and Research Centre, Sripuram, Vellore - 632055, India.

^b Environmental Molecular Microbiology Research Laboratory, Department of Biotechnology, Thiruvalluvar University, Serkadu, Vellore - 632115, India.

Authors' contributions

This work was carried out in collaboration among all authors. Authors MR and BN were involved in the study design, implementation and supervision. Authors PR and NS were involved in data collection, experimental analysis and interpretation. Author RA involved in supervision. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JABB/2022/v25i7588

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/92417>

Original Research Article

Received 04 August 2022
Accepted 08 October 2022
Published 18 October 2022

ABSTRACT

Introduction: The Rubella virus has a worldwide occurrence and congenital Rubella syndromes are widely recognized as an emerging infection in several parts of the world. Miscarriage, perinatal mortality, and stillbirth can develop in pregnancy during the first trimester. The most frequent techniques for laboratory diagnosis of Rubella virus infection are IgM and IgG-based serological detection methods. Such emerging viral and bacterial pathogen emphasizes the development of fast diagnostic devices; there is a need for enhanced and quicker methods.

Materials and Methods: Search for peptide vaccine with specific T and B-cell epitopes was identified through bioinformatics-based approaches. These were identified utilizing available Rubella virus E1 glycoprotein sequence databases. The outer-membrane glycoprotein, E1 is a target protein for the prediction of best antigens.

Results: Using BepiPred2 program, the potential B-cell epitope PFCNTPHGQLEVQVPPDPGD with high conservation among E1 glycoprotein of rubella virus

*Corresponding author: E-mail: rmageshbabu@rediffmail.com;

and the maximum surface exposed residues was identified. Using IEDB, NetMHCpan, and NetCTL programs, T-cell epitope RPVALPRAL was identified. Predicted epitopes were found to have promiscuous class-I major histocompatibility complex binding affinity to major histocompatibility complex super types, antigenicity scores, and high proteasomal cleavage. The three-dimensional modeled structures were created using I-TASSER online server for highlighting the predicted T- and B- cell epitopes.

Conclusion: The predicted T and B cell epitope could be used for the development of immunoglobulin assay and vaccine candidate peptide.

Keywords: Epitope; Rubella virus; peptide vaccine; B and T cell peptide.

1. INTRODUCTION

Rubella virus belongs to the rubivirus genus of the Togaviridae family and it is a positive sense single-strand RNA. About 25% to 50% of infections are asymptomatic [1,2]. In newborn children, CRS (Congenital Rubella Syndrome) generally causes mild illness, with visible manifestations. In infants, a red rash is usually the first sign in those who do develop symptoms. The rash usually starts on the face and then spreads to the rest of the body, lasting around three days. Worldwide more than 100000 infants are infected by CRS each year [3]. Rubella is a highly contagious disease that is spread by respiratory droplets that often manifests in babies as a low-grade fever, arthralgia, maculopapular rash, and myalgia. (WHO 2011) [4]. Infection in pregnant women in the first 12 weeks can cause foetal death or CRS, which manifests as cataract and glaucoma, neural deafness, heart disease and microcephaly in 80-90% of cases [5,6]. Studies in India indicated a high sero-prevalence of rubella infection [7]. After vaccination and infection immune response developed in the body with the development of antibodies against the structural protein, which aimed toward E1, E2 and capsid protein [8,9]. The antibodies produced against glycoprotein E1 glycoprotein continue to stay in the body for a long time, but the antibodies produce against E2 and capsid protein stay for a short time, it might differ by the accessibility of the immune system [10,11].

The present vaccine is live attenuated and available as a Measles-Mumps-Rubella (MMR) vaccine and is known to have some acute side effects [12,13]. The sero-conversion to the rubella virus is about <90% with one dose of MMR. Peptide-based vaccines with reduced adverse reactions could be developed. The present method of serological diagnosis is IgM (acute infection) or IgG (past exposure). These assays use recombinant antigens or viral antigens purified from lysate and could have low sensitivity $\leq 90\%$ and a false positive rate of 5%.

It is possible to improve the performance of these assays with peptide antigens [14]. Rubella virus infections are common for males and females, however highly risk to immuno-suppressed patients and pregnant women. Our study aimed to select T and B cell peptides of E1 glycoprotein using bioinformatics tools and the predicted peptides can be used in the development of immunoassay and vaccines. This study is focused on rubella virus vaccine development and this vaccine candidate peptide can induce an immune response against rubella virus infection.

2. MATERIALS AND METHODS

All available full-length coding sequence (n=181) of the Glycoprotein E1 gene of the rubella virus was downloaded from the NCBI database. A consensus sequence was selected from the 181 Glycoprotein E1 sequences using the CLC sequence viewer 8.0 software. Linear B cell epitopes were predicted using two different programs: BepiPred 2.0 [15] and IEDB analysis resource [16-17]. The Glycoprotein E1 gene consensus sequence was uploaded to each programme. Using the CLC Sequence viewer software, the epitope predicted by the two algorithms was evaluated for the conservancy, and a conserved B-cell epitope candidate vaccine was chosen. The glycoprotein E1 gene coding sequence 3D protein structure is stimulated by the I-TASSER web server program.

RAMPAGE program [18] was used to examine the conformation of the modeled structure and the Ramachandran plot. For backbone dihedral angles ψ versus ϕ amino acid residues, the Ramachandran plot was used to visualize energetically allowed areas in the simulated protein structure. RAMPAGE shows the proportion of residues in different places, such as the preferred region, permitted region, and outlier region. The protein becomes more stable as the number of residues in the preferred area increases. To find discontinuous epitopes, the

DiscoTope 2 program [19] was used using default prediction settings. The estimate was based on the 3D structure created by I-TASSER. The final scores are derived by adding the propensity scores of nearby residues and the surface accessibility [20]. The ElliPro software assigns a score to each anticipated epitope, which was calculated by averaging the Protrusion Index (PI) overall epitope residues.

The IEDB analysis resource software was used to predict Class-I MHC binding T-cell epitopes. The human HLA allele reference set (27 alleles) with both 9-mer and 10-mer peptide length lengths was utilized for the prediction, which was done using the IEDB-recommended prediction technique. According to the percentile rank, the epitopes were predicted. Each MHC allele has a high affinity for it, as well as its length. This indicated by a tiny numerical percentile rank (1%). As a result, available epitopes with percentile rankings below the threshold were selected for further analysis.

MHC-I binding T-cell peptide lengths of 8 - 11mer were also predicted using NetMHC 4.0 server [21], Artificial Neural Networks (ANN) is used in this system. For prediction, the HLA representatives of supertypes and other default criteria (0.5 percent Rank) were chosen. The program's strongest binders were chosen for further investigation.

TAPred [22] was also used to predict TAP-binding peptides. Binders with high and moderate affinity were chosen using a cascade SVM prediction method. NetChop 3.1 sever [23] was used to predict protoplasmic cleavage.

The IEDB analysis resource was used to predict the immunogenicity of the chosen peptides in Class I. This bioinformatics tool predicts the immunogenicity of a peptide MHC complex based on amino acid characteristics and their location within the peptide. The Vaxijen online server software was used to forecast protective antigens without regard to alignment. The physicochemical characteristics of proteins are used to classify antigens.

3. RESULTS

3.1 B-Cell Peptide Prediction

Separate epitopes were identified using two different algorithms, then listed and evaluated for conservation among the 181 sequences examined. The BepiPred 2 programme produced four peptide epitopes of different lengths. One

epitope, however, was found in the highly conserved region (PFCNTPHGGQLEVQVPPDPGD).

Among the four peptide epitopes produces by the IEDB algorithm only one peptide (PFCNTPHGGQLEVQVPPDPGD), fell within the appropriate size range. However, because the epitope was located in a very changeable area, it was deemed unsuitable.

In the I-TASSER-generated protein model, the chosen peptide PFCNTPHGGQLEVQVPPDPGD was highlighted (Fig. 1). The epitopes in the model were labeled using the Pymol software.

The estimated TM-score was 0.78 ± 0.10 , and the estimated RMSD was 6.2 ± 3.8 . The C-score was 0.49, the estimated TM-score was 0.78 ± 0.10 , and the estimated RMSD was 6.2 ± 3.8 . The C-score is a confidence score used by I-TASSER to estimate the quality of projected models. It's computed using the importance of threading template alignments and the structure assembly simulations' convergence parameters. The C-score is usually in the range of [-5, 2], with a higher C-score indicating a more confident model and vice versa. The TM-score of >0.5 suggests a model with correct topology, A random similarity is indicated by a TM-score of <0.17 . The Ramachandran plot analysis found on the Rampage server revealed that the model created by I-TASSER was close to acceptable. 82.5 percent, 11.1 percent, and 6.5 percent of residues were found in the preferred, permitted, and outlier areas, respectively (Fig. 2).

4. DISCONTINUOUS B-CELL PEPTIDE PREDICTION

Using the modeled 3D protein structure of the Glycoprotein E1 gene in PDB format, the DiscoTope 2 software predicted discontinuous epitopes, which are indicated in the modeled protein structure (Fig. 3). The positions of the detected residues are 152; 201; 209-218; 220; 226-228; 302-134. Using the Pymol software, the epitopes were highlighted in the 3D structure.

4.1 T-Cell Peptide Prediction for MHC CLASS-I

All three algorithms generated MHC-I predicted T cell peptides, which were given together with other prediction scores (Table 1). One epitope, RPVALPRAL, TAPred score was high, and the protein survived proteasomal cleavage, as

predicted by the NetChop 3.1 algorithm, as well as a high Class I Immunogenicity and Vaxijen score among the epitopes found by all three systems. HLA supertypes HLA-B*07:02 and HLA-B*39:01 exhibited a strong affinity for them. This was followed by another epitope,

RPRLRLVDA, which showed strong MHC binding, TAP prediction, NetChop 3.1-predicted proteasomal cleavage, and good antigenicity ratings. The MHC supertypes HLA-B*07:02 and HLA-B*08:01 were predicted to have a strong affinity for this epitope.



Fig. 1. 3D protein structure of E1 of Rubella virus. The linear B-cell epitope predicted by Bepipred and IEDB is highlighted in red

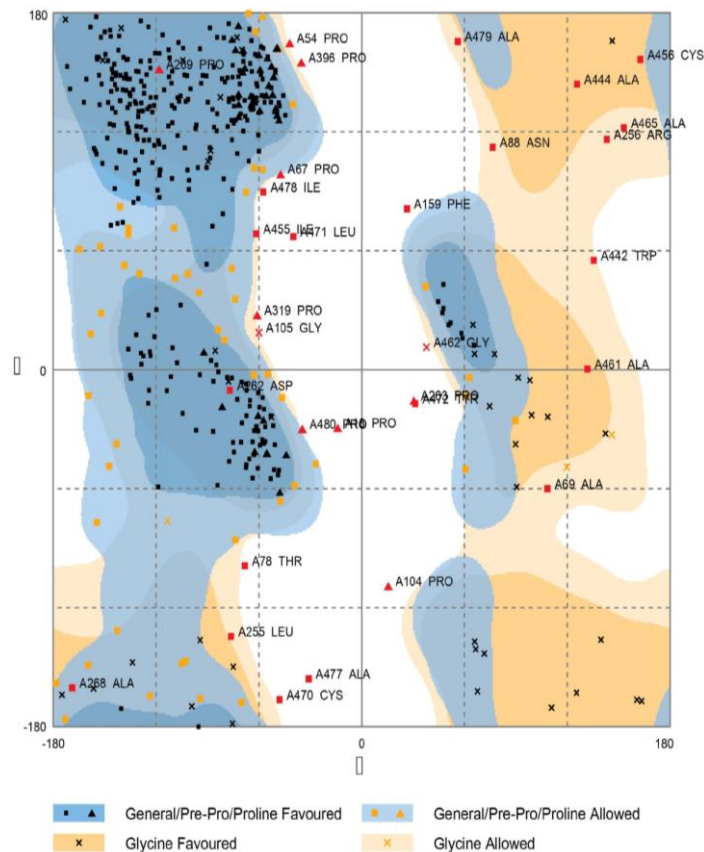


Fig. 2. Molecular characterization of Rubella virus E1 glycoprotein using RAMPAGE program with residues scores

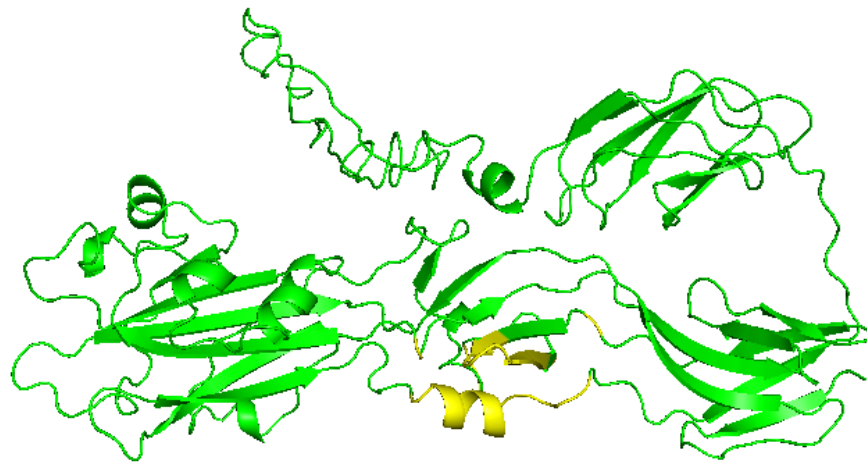


Fig. 3. Discontinuous epitopes predicted by Discotope 2 bioinformatics tool. The predicted discontinuous epitopes are highlighted in yellow

Table 1. T-cell peptides identified by three different programmes (IEDB, NetMHCpan and NETCTL)

List of epitopes	Survive Proteasomal cleavage*	TAPred score#	Vaxijen score^	Immunogenicity score
AQSFTGVVY	Yes	3.841	0.1614 (Probable NON-ANTIGEN).	0.19276
RPVALPRAL	Yes	8.363	0.6515 (Probable ANTIGEN)	0.09807
GEVWVTPVI	Yes	8.573	-0.1840 (Probable NON-ANTIGEN)	0.32779
RPRLRLVDA	Yes	8.129	1.3364 (Probable ANTIGEN).	0.0934
CTFWAVNAY	Yes	3.483	0.3954 (Probable NON-ANTIGEN).	0.35525
MSVFALASY	Yes	4.219	0.3212 (Probable NON-ANTIGEN)	0.9522
DLVEYIMNY	Yes	3.974	-0.2477 (Probable NON-ANTIGEN)	0.08385
ETRQTWAEW	Yes	5.317	1.0473 (Probable ANTIGEN)	0.23807
FHTETRTVW	Yes	6.713	0.4227 (Probable NON-ANTIGEN)	0.25675
TPERPRLRL	Yes	3.880	-0.1644 (Probable NON-ANTIGEN)	0.14338
TETRTVWQL	Yes	3.841	0.6486 (Probable ANTIGEN)	0.14338

*Proteasomal cleavage prediction by the NETchop 3.1 program

#TAPred score intermediate binding affinity

^Vaxijen score with probable antigen, antigen and non-antigenic

5. DISCUSSION

The goal of this study was to find possible epitopes that might trigger cellular and humoral immune responses that could be used to develop immunological assays and vaccines. We utilized bioinformatics tools to predict epitopes for rubella virus E1 glycoprotein. Rubella virus has three structural proteins: E1 and E2, as well as a capsid protein. The glycoprotein is responsible for cellular response recognition.

In the prediction of B-cell peptides, the Bepipred 2.0 server with an artificial neural network and the IEDB server were utilized. IEDB and NetMHC 4.0 servers based on artificial neuron networks were also used to predict T-cell epitopes. The

consensus sequence was obtained using the CLC sequence viewer 8.0 program. In addition, the physicochemical properties of B cell epitopes, including Discontinue epitopes using discotope programme, epitopes affinity prediction using TAPred, proteosomal cleavage analysis using NETCHOP, and the 3D structure was evaluated using I-TASSAR and VAXIJEN online software was used for alignment-independent prediction. Results from the above analyses were scored to establish the reliability of predicted antigenic epitopes. Rubella virus E1 glycoprotein was subjected to Bepipred 2.0 cell epitope prediction tests. There were four peptides identified, with the 20 amino acid PFCNTPHGQLEVQVPPDPGD B cell epitope from 183 to 201 being the most acceptable. For

the prediction of discontinuous B-cell epitopes, we applied Ellipro and Discotope.

The BepiPred2 algorithm predicted that the B-cell peptide epitope PFCNTPHGQLEVQVPPDPGD contained 12 exposed amino acid residues. The functional B-cell epitopes have a length of 15 - 25 residues and are surface accessible for effective antigen-antibody interactions. B-cell receptors detect surface-accessible clusters of amino acids, which can activate a cellular immune response. The generated epitopes that were within the conserved regions with good prediction score were chosen. The major portions of amino acid residues were comparable between the two programs.

We used three different programs viz. IEDB, NetMHCpan, and NetCTL for the prediction of class-I MHC binding T-cell epitopes. We found the T-cell epitope RPVALPRAL is highly conserved in rubella E1 glycoprotein. In the E1 glycoprotein, the peptide between 330 and 339 was predicted. Antigenicity of the selected epitope was predicted using VaxiJen server with default parameters, which was developed for the prediction of potent antigen and subunit vaccines with an accuracy of 70% to 85%. Antigenic peptides with a higher antigenicity score (>1.0) did not survive the program's prediction of proteasomal cleavage. However, the chosen peptide, RPVALPRAL, got a score of 0.6515, suggesting that it was antigenic to a high degree. The C-score in the ITASSER model was 0.69, while the projected TM-score was 0.78±0.10. The bioinformatics tools listed above were carefully considered in our study because different bioinformatics tools provide varied results. Several programmes were used for T and B cell peptide prediction, and only the harmonized consensus findings are given here. The overall results indicated that then epitopes identified in our study would be highly conserved in Rubella virus and would be immunogenic, and therefore could be considered as a potential candidate vaccine or suitable antigen for the development of immunodiagnosics test.

6. CONCLUSION

In this study, we used the bioinformatics approach to identify potential epitopes for B and T cells. There may be a possibility of using the epitopes of B-cell peptides as an antigen in a microplate ELISA or point-of-care test. The best possible B-cell epitope was predicted as PFCNTPHGQLEVQVPPDPGD, while the best candidate T-cell epitope was identified as

RPVALPRAL, which is conserved to rubella virus, based on the mean percent prediction probability score. These epitopes are ideal for the development of vaccines and immunoassays in the detection of antibodies to the rubella virus E1 glycoprotein gene. It could be evaluated as a vaccine candidate. E1 glycoprotein contains 80% surface exposed residues for the peptide. Patients infected with Rubella could be screened for antibodies with a vaccine candidate peptide. Identified T-cell epitopes need to be tested for pre-existing T-cell responses in individuals who have recovered from rubella virus infection. Various adjuvants can be incorporated into a vaccine to successfully treat animals and humans with an extremely multivalent vaccination. Various adjuvants can be incorporated into a vaccine to successfully treat animals and humans with an extremely multivalent vaccination.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Dominguez G, Wang CY, Frey TK. Sequence of the genome RNA of rubella virus: evidence for genetic rearrangement during togavirus evolution. *Virology*. 1990;177:225-238.
2. Battisti AJ, Yoder JD, Plevka P, Winkler DC, Prasad VM, et al. Cryo-electron tomography of rubella virus. *J Virol*. 2012;86:11078-11085.
3. George S, Viswanathan R, Sapkal GN. Molecular aspects of the teratogenesis of rubella virus. *Biol Res*. 2019;52(1):47.
4. WHO organization. Weekly epidemiological record. Rubella vaccines: WHO position paper. 2011;86:301-316.
5. Best JM. Rubella. *Semin Fetal Neonatal Med*. 2007;12:182-192.
6. Atkinson W, Wolfe S, Hamborsky J. Epidemiology and prevention of vaccine-preventable diseases. Centers for Disease Control and Prevention; 2011.
7. Muliylil DE, Singh P, Jois SK, Otiv S, Suri V, Varma V, et al. Sero-prevalence of rubella among pregnant women in India, 2017. *Vaccine*. 2018;36(52):7909-7912.
8. Mitchell LA, Tingle AJ, Décarie D, Shukin R. Identification of rubella virus T-cell epitopes recognized in anamnestic response to RA27/3 vaccine: associations

- with boost in neutralizing antibody titer. *Vaccine*. 1999;17:2356-2365.
9. Petrova EK, Dmitrieva AA, Trifonova EA, Nikitin NA, Karpova OV. The key role of rubella virus glycoproteins in the formation of immune response and perspectives on their use in the development of new recombinant vaccines. *Vaccine*. 2016;34:1006-1011.
 10. Gießauf A, Letschka T, Walder G, Dierich MP, Würzner R. A synthetic peptide ELISA for the screening of rubella virus neutralizing antibodies in order to ascertain immunity. *J Immunol Methods*. 2004;287:1-11.
 11. Seto NO, Gillam S. Expression and characterization of a soluble rubella virus E1 envelope protein. *J Med Virol*. 1994;44:192-199.
 12. Yavuz ST, Sahiner UM, Sekerel BE, Tuncer A, Kalayci O, et al. Anaphylactic reactions to measles–mumps–rubella vaccine in three children with allergies to hen’s egg and cow’s milk. *Acta Paediatr*. 2011;100:e94-e96.
 13. Sukumaran L, McNeil MM, Moro PL, Lewis PW, Winiacki SK, et al. Adverse events following measles, mumps, and rubella vaccine in adults reported to the vaccine adverse event reporting system (VAERS), 2003- 2013. *Clin Infect Dis*. 2015;60:e58-65.
 14. Binnicker MJ, Jespersen DJ, Harring JA. Multiplex detection of IgM and IgG class antibodies to *Toxoplasma gondii*, rubella virus, and cytomegalovirus using a novel multiplex flow immunoassay. *Clin Vaccine Immunol*. 2010;17(11):1734-1738.
 15. Jespersen MC, Peters B, Nielsen M, Marcatili P. BepiPred-2.0: improving sequence-based B-cell epitope prediction using conformational epitopes. *Nucleic Acids Res*; 2017.
 16. Larsen JEP, Lund O, Nielsen M. Improved method for predicting linear B-cell epitopes. *Immunome Res*. 2006;2:2.
 17. Yang J, Yan R, Roy A, Xu D, Poisson J, Zhang Y. The I-TASSER suite: protein structure and function prediction. *Nat Methods*. 2015;12:7-8.
 18. Lovell SC, Davis IW, Arendall WB 3rd, de Bakker PI, Word JM, Prisant MG, Richardson JS, Richardson DC. Structure validation by C α geometry: phi,psi and C β deviation. *Proteins*. 2003;50(3):437-50.
 19. Reliable B Cell epitope predictions: Impacts of method development and improved benchmarking jens vindahl kringelum, claus lundegaard, ole lund, and morten nielsen plos computational biology, 2012.
 20. Kringelum JV, Lundegaard C, Lund O, Nielsen M. Reliable B cell epitope predictions: impacts of method development and improved benchmarking. *PLoS Comput Biol*. 2012;8(12):e1002829.
 21. Andreatta M, Nielsen M. Gapped sequence alignment using artificial neural networks: application to the MHC class I system. *Bioinformatics*. 2016; 32(4):511-7.
 22. Bhasin M, Raghava GP. Classification of nuclear receptors based on amino acid composition and dipeptide composition. *J Biol Chem*. 2004;279(22): 23262-6.
 23. Nielsen R, Williamson S, Kim Y, Hubisz MJ, Clark AG, Bustamante C. Genomic scans for selective sweeps using SNP data. *Genome Res*. 2005;15(11):1566-1575.

© 2022 Rajendiran et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:

<https://www.sdiarticle5.com/review-history/92417>